

World Journal of Advanced Pharmaceutical and Life Sciences

Journal homepage: https://zealjournals.com/wjapls/

ISSN: 2799-0222 (Online)

(RESEARCH ARTICLE)

Check for updates

Ethnobotanical survey of medicinal plants against ophidian envenomations in the Bonginda/Bikoro Group in DR Congo

Blaise Engomba MOKEKOLA ^{1, 2}, Rombeau Tamasala NDOMBE ¹, Raphaël Kalima MWANGE ¹, Aristarque Bulambo MULONDA ¹, Florent Biduaya MUKEBA ¹, Zacharie Kusamba CHIFUNDERA ⁴ and Assumani Zabo IDRISSA ^{1, 3, *}

¹ Department of Biology, Faculty of Sciences, National Pedagogical University, Kinshasa, DR Congo. ² Higher Pedagogical Institute of MBANDAKA, Equateur DR Congo.

 ³ Interdisciplinary Research Center of the UPN Pedagogy and Public Health Research Unit, DR Congo.
⁴ Water and Forest Department, Spatial Teledetection and Telecommunication High School National Pedagogical University, Kinshasa, DR Congo.

World Journal of Advanced Pharmaceutical and Life Sciences, 2022, 02(02), 056-062

Publication history: Received on 25March 2022; revised on 10 May 2022; accepted on 12 May 2022

Article DOI: https://doi.org/10.53346/wjapls.2022.2.2.0024

Abstract

Snakebite envenomation is a neglected tropical disease causing enormous suffering, disability and premature death on all continents. This study aims to inventory the plants involved in the management of snakebite cases in rural areas.

A survey was conducted in the Bonginda groupement, Bonginda Territory, Equateur Province in DR Congo during the period from January 2019 to December 2020. Data was collected by the standardized interview method with a focus on individual interview technique. To this end, a questionnaire was administered to traditional healers reputed in the treatment of snakebites.

It shows that 17 species grouped into 14 families are used to treat cases of ophidian envenomation in Bonginda. The Fabaceae and Rubiaceae families each have 3 species. The others are monospecific. Among the organs used, leaves predominate with a score of 44.4%. They are followed by the roots (18.5%). Regarding the mode of preparation, the decoction prevails with 27.3%. It is followed by grinding (13.6%). As for the route of administration, local application (28.6%) is ahead of the oral route (23.8%) and purgation (14.3%).

Constituents based on medicinal plants are recognized as local heritage.

Keywords: Medicinal Plants; Ophidian Envenomation; Traditional Healer; Rural Environment; DR Congo

1. Introduction

Snake envenomation is a worldwide health problem and a reason for morbidity and mortality. According to WHO [1] around 5.5 million snakebites occur each year, resulting in up to 2 million envenomations, at least 100,000 deaths and around 300,000 amputations and other irreversible disabilities [1]. More than 500,000 victims of snakebite envenomation have been reported in Africa [2-5].

*Corresponding author: Assumani Zabo IDRISSA

Department of Biology, Faculty of Sciences, National Pedagogical University, Kinshasa, DR Congo.

Copyright © 2022 Author(s) retain the copyright of this article. This article is published under the terms of the Creative Commons Attribution Liscense 4.0.

In Africa, ancestors hold endogenous knowledge that they pass on to their descendants to treat tropical diseases [6]. Qualified as empirical, this endogenous knowledge is not based on any theory, but it is often used in the composition of drugs [7].

In rural Africa, the role of traditional medicine and medicinal plants in the management of snakebites is well established [8]. The majority of people bitten by snakes claim to have used traditional treatment rather than to the antivenom of modern medicine [9, 10]. Successful snakebite treatment in rural areas requires the involvement of traditional health practitioners [11].

With a view to palliating deaths due to snakebites, various authors have taken an interest in the medicinal plants used in traditional medicine against ophidian envenomations [2, 11, 12]. The use of plants against the effects of snakebite has long been recognized. Currently, various authors are giving it more scientific attention in their studies [13-16].

This study aims to inventory the plants involved in the management of snakebite cases in the Bonginda group, Bikoro Territory, Equateur Province in DR Congo.

2. Material and methods

2.1. Study framework

Reporting to the Lac Ntomba Sector, the Bonginda group constitutes the large liquid part of the territory. Administratively, the chief town of the territory is included in this grouping. Referring to the map below, the Bonginda groupement is designated by the edge. The chief town of the sector is in Moheli on the axis Mpenda - Momboyo Catholic Mission [17].



Figure 1 Map of the territory of Bikoro Source St Moulin 2005

2.2. Ethnobotanical surveys

Data collection concerns the period from January 2019 to December 2020. A questionnaire was administered to traditional healers known to treat snakebites.

The data was collected by the standardized interview method, favoring the individual interview technique. A foursection questionnaire was therefore developed:

- An Introductory Note Section;
- A Section Of The Socio-Demographic Characteristics Of The Respondent;
- A Section Of The Survey Itself;
- A Section Of Ethical Considerations

3. Results

Table 1 shows the floristic list, the vernacular names, the organs used, the methods of preparation and administration

Table 1 Floristic list, vernacular names, organs used, methods of preparation and administration

Family	Species	Vernacular name (Dialect)	Organ used	Method of preparation	Routes of administration
Annonaceae	Anonidium mannii(Oliv.) Engl. & Diels	Mondenge ya nzambe (lingala) Mobei (Lokonda, Lontomba)	Sterm bark	Splay	Scarification Local application
Caricaceae	Carica papaya L.	Paipai (lingala)	Leaf Root	Decoction	Bath
Cecropiaceae	<i>Cecropia pachystachya</i> Trécul	Sabu, Bomambo (Lomongo) Bombambo (Lonkundo) Kombokombo (Lingala) Bokombo (Lokonda , Lontomba)	Root Leaf Bud	Maceration Calcination	Oral route Massage
Commelinaceae	Palisota schweinfurthii C.B.Clarke	Zazu, Kpato, Zâzo (Ngwaka) Itele (Lontomba)	Whole plant	Splay	Scarification Local application
Euphorbiaceae	Manniophyton fulvum Müll.Arg	Mokosa (Lingala) Sambia (Lontomba)	Stem sap	Extraction	Local application
Fabaceae	Scorodophloeus zenkeri Harms	Bofili (Lomongo) Bopili (Lokonda) Mbopili (Mpama)	Root bark Leaf	Calcination	Purgation
Fabaceae	Senna occidentalis (L.) Link	Limingolanta (Lomongo) Bonungolata (Lonkundo) Bolebebonse (Lontomba) Betshobeawane(Elinga)	Leaf Pod Root	Drying Incineration of roots Cutting of leafs and pod	Incisions Local application
Fabaceae	Pentaclethra macrophylla Benth.	Ebala (Kitembo) Buadja (Lomongo) Owala (Kitetela) Boala (Lonkundo) Obala (Mpama, Kinunu) Ehili (Lokonda)	Pod	Decoction	Bath
Clusiaceae	Harungana madagascariensis Lam. ex Poir.	Botonongolo, Montone (Lonkundo) Bohili (Lontomba)	Leaf	Grinding	Local application
Meliaceae	Entandrophragma palustre Staner	Bosala (Kinunu, Mongo) Bobala (Lokonda, Lotomba)	Stem bark	Decoction	Purgation Inhalation

Phyllanthaceae	HymenocardiaulmoidesOliv.	Ikengeleke (Lokonda) Ikengelek (Lonkundo) Eyanze (Lontomba)	Leaf	Decoction	Oral route
Rubiaceae	Feretia apodanthera Del.	Ilongia (Ekonda , Lontomba)	Leaf	Pounding Maceration	Oral route
Rubiaceae	<i>Morinda morindoides</i> (Baker) Milne-Redh	Kongobololo (Lingala, Lokonda) Kong'ololo (Lonkundo) Ngongabololo (Mpama)	Leaf	Decoction	Oral route
Rubiaceae	Ophiorrhiza mungos Linn	Nsambiilokay (Lontomba)	Leaf	Chewing	Oral route
Salicaceae	Oncoba welwitschii Oliv	Ehake (Lotomba, Lokonda)	Leaf Root	Decoction Filtration	Purgation
Sapotaceae	Synsepalum dulcificum Schum.) Baill.	Bomonga, Mpunga (Lontomba) Bompunga (Lokonda) Munga (Ngwaka) Mofunga (Elinga)	Stem Leaf Root	Grinding	Local application
Solanaceae	Nicotiana tabacum L.	Ikaya li tumbaco (Kisengele) Mongolo (Lontomba) Likaya (Mpama) Tumbaco (Lingala)	Leaf	Grinding	Local application

The analysis of Table 1 reveals that in Bonginda, the anti-venom floristic list includes 17 species grouped into 14 families. The Fabaceae and Rubiaceae families each have 3 species. The others are monospecific.

Figure 2 presents the analysis of the organs used, the methods of preparation and the routes of administration



Figure 2 Organs used, Method of preparation and Routes of administration

3.1. Organs used

The leaves predominate with the score of 44.4%. They are followed by the roots (18.5%). Root bark, stem bark and pod each account for 7.4%. They precede the buds, the whole plant, the stem sap and the stem which each represents 3.7%.

3.2. Method of preparation

The decoction takes precedence with 27.3% followed by grinding (13.6%). Calcination, maceration and pulverization each intervene with 9.1%.

3.3. Route of administration

Local application (28.6) is ahead of the oral route (23.8%) and purgation (14.3%). The local bath (9.5%) and the scarification (9.5%) predominate over the incision, the inhalation, and the massage which intervene each with 4.8%.

4. Discussion

The treatment of snakebites by plants is of interest to several researchers around the world. In Bonginda, most cases of ophidian envenomation call on medicinal plants through traditional healers. A list of 17 species grouped into 14 families has just been drawn up. The Fabaceae and Rubiaceae families each have 3 species.

- Fabaceae: Scorodophloeus zenkeri Harms; Senna occidentalis (L.) Link; Pentaclethra macrophylla Benth.
- Rubiaceae: Feretia apodanthera Del.; Morinda morindoides (Baker) Milne-Redh; Ophiorrhiza mungos Linn.

Ngbolua *et al.*[9] note that 17 medicinal plants belonging to 16 botanical families are used against snakebites in forest areas of DR Congo. The families Leguminosae and Araceae each have 2 species.

- Leguminoseae: Senna occidentalis (L.) Link, Piptadeniastrum africanum (Hook.f.) Brenan
- Araceae: Anchomanes giganteus Engl. ; Cercestis congoensis Engl.

In Uganda, a literature review carried out in multidisciplinary databases revealed that 77 plant species belonging to 65 genera and 42 families are used for the treatment of snakebites in Uganda [12]). The majority of these species belong to the Fabaceae (31%), Euphorbiaceae (14%), Asteraceae (12%), Amaryllidaceae (10%) and Solanaceae (10%) families.

In Kenya, Omara[2] identified 54 species grouped into 27 families. The predominant families consisted of Fabaceae and Malvaceae, each with 5 species.

In Senegal, Sow [18] carried out a survey at three sites in three regions: Dakar, Kaolack and Kédougou. In total, he recorded two families of plants both among healers, traditional healers and herbalists:

- Annonaceae: Annona chrysophylla Boj., A. senegalensis var. chrysophylla (Boj.) R. Sillans, A. senegalensis var latifolia Oliv., A. arenaria Thonn;
- Polygalaceae: Securidaca longipedunculata Fresen.

In Mali, Bah *et al.*[11] identified 13 species grouped into 11 families. The Rubiaceae and Fabaceae families are predominant:

- Fabaceae: Acacia albida Delile Balazan; Piliostigma reticulatum (DC.) Hochst; Tamarindus indica L.
- Rubiaceae: Crossopteryx febrifuga (Afzel) Benth. Gardenia ternifolia Schumach. & Thonn

Recognition of the anti-venom properties of certain species goes beyond the limits of the DR Congo. Indeed, *Senna siamea* is also included in the inventory carried out in Kenya [2]. The same is true for *Nicotiana tabacum, Senna occidentalis, Carica papaya* in Uganda [12] and *Ophiorrhiza mungos* Linn. in India [19].

Molander *et al.*[20], tested 226 extracts of 94 species used to treat snakebites from three countries: Mali, D.R. Congo and South Africa. The Anacardiaceae and Malvaceae families had the highest number of active species. Of the 226 plant extracts tested, 41 plant species showed more than 90% inhibition in one or more tests. Inhibition is attributed mainly to polyphenols such as tannins in most extracts. Inhibition of digestive enzymes by tannins may be beneficial in the treatment of local tissue damage caused by snake venom and inhibition of the "spreading factor" The enzyme hyaluronidase may delay the onset of systemic effects, thereby improving the rate of survival.

Among the 41 plant species, five plant extracts were active in the hyaluronidase enzyme assay and one extract was active in the protease assay after removal of tannins suggesting that they may contain a more specific inhibitor. The five plants are widely distributed alongside *Bitis arietans* and *Naja nigricollis* is a potential inhibitor accessible to everyone in sub-Saharan Africa [20].

Many Indian herbal medicines are recommended for the treatment of snakebites. Several laboratory studies have demonstrated the anti-venom effects of plant extracts. Let us mention in particular the cases of:

- Methanolic root extracts of *Vitex negundo* Linn. and *Emblica officinalis* Gaertn. [13]
- Seed extract of *Strychnos nuxvomica* Linn [21];
- Leaf extracts of Asystasia gangetica (L) and Newbouldia leavis (P. Beauv) [22]
- Plant extracts of Andrographis paniculata and Aristolochia indica [23]
- Extract of Eclipta prostrata [24]
- Extract root of *Tabernaemontana alternifolia* [25].

5. Conclusion

Given the prevalence of snakebite cases in the equatorial forest and the resulting consequences, the present study aimed to enhance endogenous knowledge on the treatment of ophidian envenomation in Bonginda, Territory of Bikoro Province of Ecuador in DR Congo.

The field survey revealed that the anti-venom floristic list includes 17 species grouped into 14 families. The Fabaceae and Rubiaceae families each have 3 species.

- Fabaceae: Scorodophloeus zenkeri Harms; Senna occidentalis (L.) Link; Pentaclethra macrophylla Benth.
- Rubiaceae: Feretia apodanthera Del. ; Morinda morindoides (Baker) Milne-Redh; Ophiorrhiza mungos Linn

The reputation of the anti-venom properties of certain species goes beyond the African continent. Such is the case of *Ophiorrhiza mungos* Linn. prescribed in India against the same pathology

Compliance with ethical standards

Acknowledgments

We thank all the stakeholders who participated in this study. Our thanks go more particularly to the traditional healers for their agreement.

Disclosure of conflict of interest

There is no conflict of interest be the authors of this manuscript.

References

- [1] World Health Organization, WHO guidelines for the production, control and regulation of snake antivenom immunoglobulins. *Geneva: WHO*, 2010, vol. 134.
- [2] Omara T. Plants Used in Antivenom Therapy in Rural Kenya: Ethnobotany and Future Perspectives *Journal of Toxicology*. 2020 Vol 2020
- [3] Chippaux JP. Snake-bites: appraisal of the global situation. Bull World Health Organ. 1998;76:515–24.
- [4] Warrell DA, Arnett C. The importance of bites by the saw-scaled or carpet viper (Echiscarinatus): epidemiological studies in Nigeria and a review of the world literature. *Acta Trop.* 1976;33:307–41.
- [5] Theakston RDG. Warrell DA, Griffiths E. Report of a WHO workshop on the standardization and control of antivenoms. *Toxicon.* 2003;41:541–57.
- [6] Zabo I.A, Tamasala N. and Diakedika L. Phytochemical study of medicinal plants used against buruli ulcer by Ntandu people in Kongo Central, DRC *Tropical Plant Research.* 2019; 6(1): 49–53.

- [7] Zerbo P., Rasolodimby; J.M., Ouedraogo O.N., et Van Damme P. Plantes médicinales et pratiques médicales au Burkina Faso: cas des Sanan. *Bois & Forets Des Tropiques*, 2011, vol. 307, p. 41-53.
- [8] Mbuta, K.K., Mwima, K., Bitengeli, I. Y.O, Kavuna M. Mabdabga M.K. Izamajole N. et Lody D. Plantes médicinales de traditions province de l'equateur-rd congo. *Institut de Recherche en Sciences de la Santé, Kinshasa, RD Congo*, 2012.
- [9] Koto-te-nyiwa N., Nagahuedi J. M.S., Sodi, Masengo, C., Ashande, R. D. D., Mpiana, P. T., & Virima, M. Synthèse bibliographique sur les serpents et les plantes médicinales utilisées en médecine traditionnelle contre les envenimations ophidiennes. *IJAR*, 2021, vol. 7, no 4, p. 305-314.
- [10] Mokekola BE, Idrissa AZ, Chifundera ZK, Ndombe RT, Mwange RK, Mulonda AB, Mukeba FB. Evaluation of snakebites and their treatment in the Bonginda/Bikoro DR Congo. World Journal of Biology Pharmacy and Health Sciences, 2022, 10(01), 039–045.
- [11] Bah S, Maiga S, Dabo A, Diarra S, Sacko M, Sanogo R, Diallo D. Utilisation des plantes médicinales dans le traitement des morsures de serpent dans le cercle de Kolokani au Mali. *Mali Santé Publique* 2011, p. 49-52.
- [12] Omara T, Kagoya S, Openy A, Omute T, Ssebulime S, Kiplagat KM, Bongomin O. Antivenin plants used for treatment of snakebites in Uganda: ethnobotanical reports and pharmacological evidences *Tropical Medicine and Health*. 2020; 48:6.
- [13] Alam MI, Auddy B, Gomes A. Isolation purification and partial characterization of Viper venom neutralizing factor from the root extract of Indian medicinal plant Hemidesmus indicus R. *British Toxicon*. 1994; 32: 155–1557.
- [14] Alam MI, Gomes A. Snake venom neutralization by Indian medicinal plants (Vitex negundo and Emblica officinalis) root extracts. *Journal of Ethnopharmacology*. 2003; 86: 75–80.
- [15] Chatterjee I, Chakravarty AK, Gomes A. Daboia russellii and Naja kaouthia venom neutralization by lupeol acetate isolated from the root extract of Indian sarsaparilla Hemidesmus indicus R.Br. Journal of Ethnopharmacology. 2006; 106: 38–43.
- [16] Mors WB, do Nascimento MC, Pereira BMR, Pereira NA. Plant natural products active against snake bite-the molecular approach. *Phytochemistry.* 2000; 55: 627–642.
- [17] De Saint Moulin L. Atlas de l'organisation administrative de la République Démocratique du Congo. Cepas. 2005.
- [18] Sow, P. G. Enquête ethnobotanique et ethnopharmacologique des plantes médicinales de la pharmacopée sénégalaise dans le traitement des morsures de serpents. *Le Pharmacien Hospitalier et Clinicien*, 2012, 47,(1) : 37-41.
- [19] Vaidya SM, Singh AR, Patel VG, Khan NA, Yewale RP,Kale MK. A review on herbs against snake *venom Journal of Pharmacognosy and Phytochemistry*. 2018; SP6: 05-09.
- [20] Molander M, Nielsen L, Søgaard S, Staerk D, Rønsted N, Diallo D, Chifundera KZ, StadenJv, JägerAK. Hyaluronidase, phospholipase A2 and protease inhibitory activity of plants used in traditional treatment of snakebite-induced tissue necrosis in Mali, DR Congo and South Africa *Journal of Ethnopharmacology*. 2014; 157:171–180.
- [21] Chatterjee I, Chakravarty K, Gomes A. Antisnake venom activity seed extract of Strychnosnux vomica Linn. *Indian Journal of Experimental Biology*.May 2004; 42: 468-475
- [22] Enenebeaku CK, Umerie SC, Nwankwo MU, Enenebeaku UE. Anti-Snake venom Activities of the leaf extracts of Asystasia gangetica (L) and Newbouldia leavis (P. Beauv) *WNOFNS*. 2018; 16: 33-41.
- [23] Meenatchisundaram S, Parameswari G, Michael A. Studies on antivenom activity of Andrographis paniculata and Aristolochia indica plant extracts against Daboia russelli venom by in vivo and in vitro methods *Indian Journal of Science and Technology*. Mar. 2009; 2(4): 76-79.
- [24] Pithayanukul P, Laovachirasuwan S, Bavovada R, Pakmanee N, Suttisri R. Anti-venom potential of butanolic extract of Eclipta prostrata against Malayan pit viper venom. *Journal of Ethnopharmacology*. 2004; 90:347–352.
- [25] Vineetha MS, Bhavya J,More SS. Inhibition of pharmacological and toxic effects of Echis carinatus venom by Tabernaemontana alternifolia root extract. *Indian Journal of Natural Products and Resources*. March 2019; 10(1): 48-58.