

Ultradian and hyperdian rhythms determine the output of transmitters from brain neurons

Athineos Philippu *

Department of Pharmacology and Toxicology, University of Innsbruck, Austria.

World Journal of Advanced Pharmaceutical and Medical Research, 2024, 06(01), 023–028

Publication history: Received on 29 December 2023; revised on 07 February 2024; accepted on 10 February 2024

Article DOI: <https://doi.org/10.53346/wjapmr.2024.6.1.0032>

Abstract

Using the push-pull superfusion technique for determining the in vivo release the neurotransmitters catecholamines, histamine, GABA and the neuromodulator nitric oxide in distinct brain areas revealed that they are released according to ultradian rhythms with frequencies of 135-36 min per cycle and hyperdian rhythms with frequencies of less than 30 min (24 -10 min) per cycle. The electrical activity of EEG delta and theta waves also varies rhythmically. The pacemaker of the EEG and histamine rhythm is located in the hypothalamus. Simultaneous recordings of EEG and histamine release revealed that high neuronal activity of EEG coincides with low histamine release rates and vice versa. The significance of these rhythms for brain function and modulation of the activity of peripheral organs by the brain is discussed.

Keywords: Brain; Push-pull superfusion; Neurotransmitters; Neuromodulators; Nitric oxide; EEG; Ultradian rhythms; Hyperdian rhythms

1. Introduction

Use of the push-pull superfusion technique (PPST) makes it possible to determine in the synapse continuously for several hours and in short time periods minute concentrations of neurotransmitter and neuromodulators released in distinct brain areas from their neurons. EEG waves may be simultaneously recorded [1-4]. Our studies revealed that in all brain areas investigated the spontaneous release rates of neurotransmitters and nitric oxide are not constant but vary according to ultradian rhythms and rhythms shorter than the ultradians, the hyperdian rhythms observed under particular experimental conditions.

2. Results and discussion

2.1. Ultradian rhythms

2.1.1. Catecholamines

Superfusion of the posterior hypothalamus of anaesthetized cats through the PPS over 6 hours and collection of the superfusates in time periods of 15 min revealed that the release rates of the catecholamines dopamine, noradrenaline and adrenaline varied from sample to sample. When the peak release rates of each experiment were synchronized, 5 peaks with high release rates in 6 hours appeared (Fig. 1). Thus, in this region of the cat brain the release rates of all three catecholamines are not constant but fluctuate according to an ultradian rhythm with a frequency of 70 min per cycle. Similar frequencies were observed in the anterior rat hypothalamus [5, 6]. Findings are summarized in Table 1.

* Corresponding author: Athineos Philippu

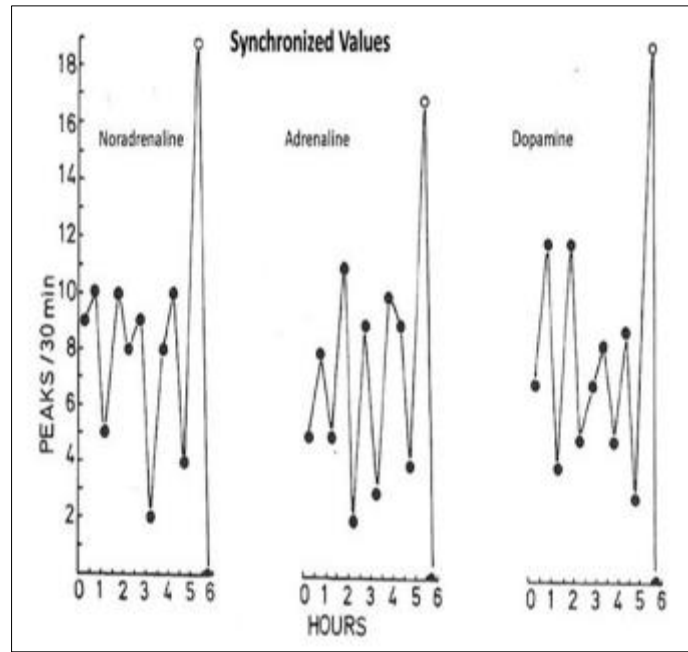


Figure 1 Spontaneous release of endogenous catecholamines in the cat posterior hypothalamus after synchronization of values shown in Fig. 1. Samples were continuously collected in 15 min periods. The last peak of high rate of release was taken as the antepenultimate sample (5.5 h) and the other samples were re-arranged accordingly. After this synchronization five peaks of high release became visible in each experiment. For differences between phases of high rate of release (5 phases, number of observations 85–95 for each catecholamine) and phases of low rates of release (5 phases, number of observations 85–95 for each catecholamine): noradrenaline, adrenaline, dopamine $P < 0.001$ (Philippu et al. 1979). Reproduced with kind permission of Springer-Verlag

Table 1 Ultradian and hyperdian rhythms of NO, neurotransmitter and EEG in various brain regions

	Animal Species	NAC	PH	AH	NTS	LC	MAN	MB
NO	an. rat	24*						
EEG	an. rat		100					
DA	an. cat		70	70	60 10*	37		
NA	an. cat		70	70	60 10*	52		
A	an. cat		70	70	60 10*	36		
HI	an. cat	60	60				135 9.5*	90 18*
HI	an. Rat co. Rat		115 90					
HI	co. rabbit	70	70	70				
GABA	co. rat		65					
GABA	an.cat	70	70					

Ultradian and hyperdian rhythms in min. Samples were collected in time periods of 10 min or less than 2.5* min. NAC nucleus accumbens, PH posterior hypothalamus, AH anterior hypothalamus, NTS nucleus of the solitary tract, LC locus coeruleus, MAN medial amygdaloid nucleus, MB mammillary body, NO nitric oxide, DA dopamine, NA noradrenaline. A adrenaline, HI histamine. GLU glutamate, ARG arginine, Tau taurine, an. anaesthetized, co. conscious. For references see text. Modified from [7].

Most of the noradrenergic and adrenergic neurons ending in the hypothalamus arrive from the locus coeruleus [8, 9], which also contains catecholaminergic nerve endings [9, 10]. Investigation of the catecholamine release in the locus coeruleus using the PPST showed that in this area the release of dopamine, noradrenaline and adrenaline also oscillates with a frequency of 37, 52 and 36 min per cycle, respectively [11]. The dissimilar ultradian rhythms of noradrenaline from those of dopamine and adrenaline point to the release of the amines from different neuronal sites (Table 1).

Similar ultradian rhythms were found in the nucleus of the solitary tract [12].

2.1.2. Amino acids

Similar to catecholamines, the inhibitory amino acid GABA is released in the posterior hypothalamus of conscious and anaesthetized rats according to an ultradian rhythm with a frequency of 65 min and 70 min per cycle, respectively [13]. Hypothalamic GABA originates to a great part from GABAergic neurons, because its release is enhanced on superfusion with potassium rich artificial cerebrospinal fluid and greatly reduced on hypothalamic superfusion with the neurotoxin tetrodotoxin [14].

2.1.3. Histamine

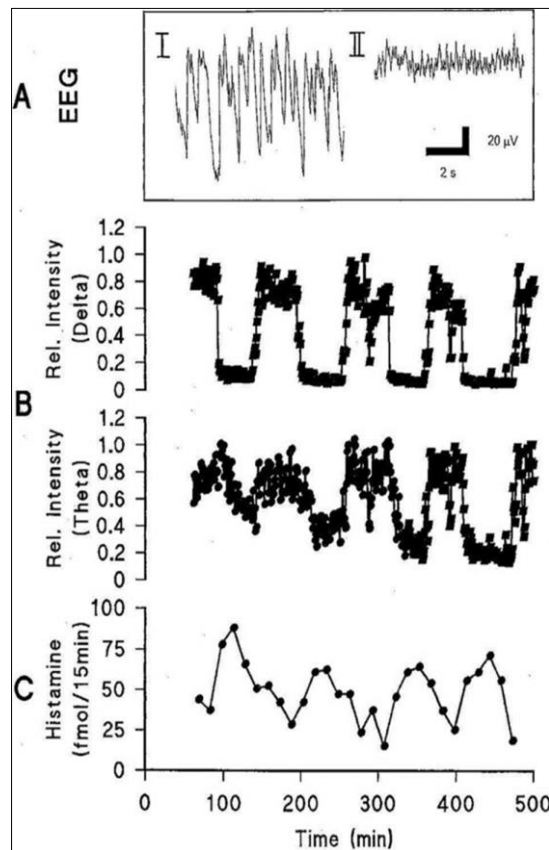


Figure 2 Two states of electrical activity in the posterior hypothalamus of the anesthetized rat. I Period of high electrical activity, II period of low electrical activity. Calibrations are indicated by the bars. Representative experiment showing the relative power the EEG 1-min mean power values in the delta (1.25–4.50 Hz) and theta (4.75–6.75 Hz) frequency waves and the release rates of histamine. Ordinates: B Relative power intensities of the delta and theta frequency waves, C Release of histamine as fmol/15 min, abscissa: time in min. Histamine: means of 7 experiments \pm S.E.M. Correlation coefficients: histamine versus delta frequency wave $R = -0.5797$ ($P < 0.001$), histamine versus theta frequency wave $R = 0.5106$ ($P < 0.01$). Reproduced from Prast et al. 1997 with kind permission of Springer-Verlag

The cell bodies of histaminergic neurons ending in the posterior hypothalamus are located in the tuberomammillary nucleus [15]. In the cat posterior hypothalamus the release of histamine oscillates with a frequency of one cycle per 60 min. In the hypothalamus of conscious and anaesthetized rats the frequency is 90 and 115 min, respectively, while in the posterior and anterior hypothalamus of the conscious rabbit (Table 1), the frequency amounts to is 70 min [16].

In the median amygdaloid nucleus and the mamillary body histamine release rates fluctuate with frequencies of one cycle every 135 min and 90 min, respectively (Table 1). The bilateral electrocoagulation of the suprachiasmatic nucleus decreased the release of histamine in the mamillary body without influencing it in the amygdaloid nucleus. indicating that in the mamillary body the release of histamine is regulated by excitatory neurons arising in the suprachiasmatic nucleus [17].

Interestingly, the long frequencies of histamine release are similar to those of EEG power (Table 1). The simultaneous determination of histamine release rates and EEG recordings has shown that the low histamine release rates coincide timely with high neuronal activity of EEG (Figure 2) and vice versa [18]. The close relationship between the fluctuations of histamine release and EEG power is underlined through the observation that agonists and antagonists of histamine receptors influence the EEG rhythm (19).

The pacemaker for the EEG power oscillations is obviously in the rostral arcuate nucleus and median eminence, because the oscillations disappear after electrocoagulation of these areas [7, 20, 21]. Possibly this pacemaker is also responsible for the oscillations of histamine release rates in the hypothalamus [14, 18, 20, 22, 23].

2.2. Hyperdian rhythms

As already mentioned, in all investigations described above the superfusates of the various brain regions were collected in time periods of 10 or 15 min. To be able to collect them in time periods down to 10 s, the speed of superfusion was slightly increased so as to increase the volume per sample necessary for the biochemical analyses. This is only possible when PPST is used and makes it feasible to investigate whether rhythms exist which are more rapid than the described ultradian rhythms. These very rapid rhythms are named hyperdian rhythms.

2.2.1. Catecholamines

In the cat nucleus of the solitary tract collection of the samples in short time periods of 2.5 min or less revealed that all three catecholamines [24] are additionally released according to a rapid hyperdian rhythm with the frequency of 10 min per cycle (Table 1).

2.2.2. Histamine

Continuous collection of the superfusate in time periods of 2 min instead of 20 min as above disclosed that in the amygdala and mamillary body of the anaesthetized cat [25] histamine is additionally released according to hyperdian rhythms with frequencies of 19.5 and 18 min, respectively (Table 1).

2.2.3. Nitric Oxide

Nitric oxide is considered to be the universal modulator of brain function [26 - 28]. To determine the release of nitric oxide under real-time conditions, a sensitive and specific amperometric sensor was inserted into the PPC. In the nucleus accumbens of rats (Table 1) the values of nitric oxide also oscillate with a frequency of one cycle per 24 min [28, 29]. This finding seems to be of particular interest is one considers the modulation of brain activity by nitric oxide [27,28]. Surprisingly is the similarity of the duration of the hyperdian rhythm in the release of nitric oxide and the duration of the hyperdian rhythm that modulates the release of histamine (see above).

3. Conclusion

Neurotransmitters and neuromodulators like nitric oxide are released in the brain according to ultradian and hyperdian rhythms. The rhythmic changes of the transmitter concentrations in the synaptic cleft influence the activity of peripheral systems, such as the cardiovascular system. Further investigations will clarify the importance of these biorhythms for the activity of central and peripheral neurons and their mutual interferences. A pacemaker located in the rostral arcuate nucleus and median eminence is responsible for the EEG power oscillations in the hypothalamus because they disappear after electrocoagulation of these areas. Possibly this pacemaker also generates the oscillations of histamine release rates in the hypothalamus.

Compliance with ethical standards

Disclosure of conflict of interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Statement of ethical approval

All animal care and experimental protocols were approved by the Commission for Animal Experiments, Federal Ministry of Science, Research and Art, Austria. All procedures used were as humane as possible. The ARRIVE guidelines for reporting experiments involving animals have been followed (Kilkenny et al., 2010; McGrath et al., 2010).

References

- [1] Philippu A. Use of push-pull cannulae to determine the release of endogenous neurotransmitters in distinct brain areas of anaesthetized and freely moving animals. In: Ed Marsden CA, . Measurement of Neurotransmitter Release. Chichester: John Wiley 1984; p. 3-37.
- [2] Hornick A, Philippu Principles of stereotaxy in small animals. In: Ed Philippu A. In vivo Neuropharmacology and Ultradian and Hyperdian Transmitter Release Journal of Advanced Neuroscience Research, 2023, Vol. 10, No. 1 7Neurophysiology. USA: Springer 2016; p. 3-14.https://doi.org/10.1007/978-1-4939-6490-1_1
- [3] Kraus MM, Philippu A. Use of push-pull superfusion technique for identifying neurotransmitters involved in brain functions, achievements and perspectives. Current Neuropharmacology 2015 Nov; 13 (6): 819-29.<https://doi.org/10.2174/1570159X1366615072233149>
- [4] Philippu A, Kraus MM. Push-pull superfusion: A technique for investigating involvement of neurotransmitters in brain function. In: Philippu A, editor. In vivo Neuropharmacology and Neurophysiology. USA: Springer 2016; p. 209-36.https://doi.org/10.1007/978-1-4939-6490-1_10
- [5] Philippu A, Dietl H, Sinha JN. In vivo release of endogenous catecholamines in the hypothalamus. Naunyn-Schmiedeberg's Arch Pharmacol 1979 Dec; 308 (1): 137-46. <https://doi.org/10.1007/BF00499055>
- [6] Dietl H, Prast H, Philippu, A. Pulsatile release of catecholamines in the hypothalamus of conscious rats. Naunyn-Schmiedeberg's Arch Pharmacol 1993 Apr; 347 (2): 28-33. <https://doi.org/10.1007/BF00168768>
- [7] Philippu A. Neurotransmitters are released in brain areas according to ultradian rhythms: Coincidence with ultradian oscillations of EEG waves. J Chem Neuroanat 2019 Mar 96 (3): 66-72. <https://doi.org/10.1016/j.jchemneu.2018.12.007>
- [8] Dahlström A, Fuxe K. Evidence for the existence of monoamine-containing neurons in the central nervous system. I. Demonstration of monoamines in the cell bodies of brainstem neurons. Acta Physiol Scand 1964; [Suppl 232]62: 1-55.
- [9] Sakai K. Physiological properties and afferent connections of the locus coeruleus and Adjacent Tegmental neurons involved in the generation of paradoxical sleep in the cat. In: Eds Banrnes CD, Pompeiano P. Neurobiology of the Locus Coeruleus. Amsterdam: Elsevier 1991; p. 31-45.[https://doi.org/10.1016/S0079-6123\(08\)63798-X](https://doi.org/10.1016/S0079-6123(08)63798-X)
- [10] Sakai K, Yoshimoto Y, Luppi PH, El Mansari M, Salvetti D, Jouvett, M. Lower brainstem afferents to the cat posterior hypothalamus double-labelling study. Brain Res Bull 1990 Mar; 24 (3): 437-55.[https://doi.org/10.1016/0361-9230\(90\)90098-K](https://doi.org/10.1016/0361-9230(90)90098-K)
- [11] Singewald N, Schneider C, Pfitscher A, Philippu A. In vivo release of catecholamines in the locus coeruleus. Naunyn-Schmiedeberg's Arch Pharmacol 1994 Oct; 350 (4): 339-45.<https://doi.org/10.1007/BF00178948>
- [12] Reis DJ, Weinbren M, Corvelli A. A circadian rhythm of norepinephrine regionally in cat brain: its relationship to environmental lighting and the regional diurnal variations in brain serotonin. J Pharmacol Exp Ther 1968 Nov; 164 (1): 135-45.
- [13] Dietl H, Philippu A. In vivo release of endogenous gamma aminobutyric acid in the cat hypothalamus. Naunyn-Schmiedeberg's Arch Pharmacol 1979 Aug 308 (2): 143-7. <https://doi.org/10.1007/BF00499056>

- [14] Singewald N, Guo L, Philippu A. Release of endogenous GABA in the posterior hypothalamus of the conscious rat; effects of drugs and experimentally induced blood pressure changes. *Naunyn-Schmiedeberg's Arch Pharmacol* 1993 Apr 347 (4): 402-6.<https://doi.org/10.1007/BF00165390>
- [15] Watanabe T, Taguchi Y, Shiosaka S, Tanaka J, Kubota, H, Terano Y et al. Distribution of the histaminergic neuron system in the central nervous system of rats; a fluorescent immunohistochemical analysis with histidine decarboxylase as a marker. *Brain Res* 1984 Mar 295 (1): 13-25.[https://doi.org/10.1016/0006-8993\(84\)90811-4](https://doi.org/10.1016/0006-8993(84)90811-4)
- [16] Prast H, Dietl H, Philippu A. Pulsatile release of histamine in the hypothalamus of conscious rats. *J Auto Nerv Syst* 1992 Jun 39 (2): 105-110. [https://doi.org/10.1016/0165-1838\(92\)90050-Q](https://doi.org/10.1016/0165-1838(92)90050-Q)
- [17] Prast H, Saxer A, Philippu A. Pattern of in vivo release of endogenous histamine in the mamillary body and the amygdala. *Naunyn-Schmiedeberg's Arch Pharmacol* 1988 Jan 337 (1): 53-7. <https://doi.org/10.1007/BF00169476>
- [18] Prast H, Grass K, Philippu A. The ultradian EEG rhythm coincides temporally with the ultradian rhythm of histamine release in the posterior hypothalamus. *Naunyn-Schmiedeberg's Arch Pharmacol* 1997 Oct 356 (4): 526-8.<https://doi.org/10.1007/PL00005086>
- [19] Prast H, Grass K, Philippu A. Influence of histamine receptor agonists and antagonists on ultradian rhythm of EEG in the posterior hypothalamus of the rat. *Neurosci Lett* 1996 Sep 216 (1):21-24.[https://doi.org/10.1016/0304-3940\(96\)12992-X](https://doi.org/10.1016/0304-3940(96)12992-X)
- [20] Grass K, Prast H, Philippu A. Ultradian rhythm in the delta and theta frequency bands of the EEG in the posterior hypothalamus of the rat. *Neurosci Lett* 1995 May 191 (3):161-4.[https://doi.org/10.1016/0304-3940\(95\)11581-G](https://doi.org/10.1016/0304-3940(95)11581-G)
- [21] Grass K, Prast H, Philippu A. Influence of mediobasal hypothalamic lesion and catecholamine receptor antagonists n ultradian rhythm of EEG in the posterior hypothalamus of the rat. *Neurosci Lett* 1996 Mar 207 (2): 93-6.[https://doi.org/10.1016/0304-3940\(96\)12494-0](https://doi.org/10.1016/0304-3940(96)12494-0)
- [22] Philippu A, Hanesch U, Hagen R, Robinson R. Release of endogenous histamine in the hypothalamus of anaesthetized cats and conscious, freely moving rabbits. *Naunyn-Schmiedeberg's Arch Pharmacol* 1982 Dec 321 /4): 282-6.<https://doi.org/10.1007/BF00498514>
- [23] Shannahoff-Khalsa D, Philippu A. A Model for the Hypothalamic Regulation of the Lateralized Ultradian Rhythms of the Autonomic and Central Nervous Systems and Psychophysiological States. *Frontiers in Psychiatry* 2023 (in press).
- [24] Lanzinger I, Kobilansky C, Philippu A. Pattern of catecholamine release in the nucleus tractus solitarii of the cat. *Naunyn-Schmiedeberg's Arch Pharmacol* 1989 Mar 339 (3):298-301.<https://doi.org/10.1007/BF00173581>
- [25] Prast H, Saxer A, Philippu A. Pattern of in vivo release of endogenous histamine in the mamillary body and the amygdala. *Naunyn-Schmiedeberg's Arch Pharmacol* 1988 Jan 337 (1): 53-7.<https://doi.org/10.1007/BF00169476>
- [26] Prast H, Tran MH, Fischer H, Philippu A. Nitric oxide-induced release of acetylcholine in the nucleus accumbens: Role of cyclic GMP, glutamate, and GABA. *J Neurochem* 1998 Jul 71 (1): 266-73.<https://doi.org/10.1046/j.1471-4159.1998.71010266.x>
- [27] Prast H, Philippu A. Nitric oxide as modulator of neuronal function. *Prog Neurobiol* 2001 May 64 (1):51-68.[https://doi.org/10.1016/S0301-0082\(00\)00044-7](https://doi.org/10.1016/S0301-0082(00)00044-7)
- [28] Philippu A Nitric Oxide: A universal modulator of brain function. *Curr Med Chem* 2016 Dec 59 (12): 2643-52.<https://doi.org/10.2174/0929867323666160627120408>
- [29] Prast H, Hornick A, Kraus MM, Philippu A. Origin of endogenous nitric oxide released in the nucleus accumbens under real-time in vivo conditions. *Life Sci* 2015 Aug 134 (1): 79-84.<https://doi.org/10.1016/j.lfs.2015.04.021>