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In vitro cytotoxic, thrombolytic, anthelmintic and antioxidant activities of *Litsea monopetala*: A medicinal plant

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Abstract

The present study was conducted to assess the cytotoxicity, thrombolytic, anthelmintic and antioxidant activity of mehanolic extract of *Litsea monopetala* (Family: Lauraceae) leaves in laboratory using in vitro methods. Cytotoxicity test was done by brine shrimp lethality bioassay where the extract concentration was 400, 200, 100, 50, 25, 12.5, 6.25, 3.125, 1.5625, 0.78125 (µg/ml). *In vitro* thrombolytic activity of *Litsea monopetala* was performed by clot lysis method using extract concentration 2.5, 5, 10 and 20 (mg/mL) in saline water. Anthelmintic activity test was done by using adult earthworms where 10, 20, 40, 60, 80 (mg/ml) extract concentration were used. Finally antioxidant activity was determined by total phenolic content determination using Folin-Ciocalteu reagent. The *Litsea monopetala* extract showed cytotoxic activity against brine shrimp nauplii and LC50 value was 41.05(µg/ml) and the investigated thrombolytic activity in our research was 9.52, 9.49, 13.64 and 17.50 % respectively as % of clot lysis. The paralysis time were at 76.75 min, 60 min, 51.75 min, 44.5 min and 64.5 min and death were at 90.50min, 63.75min, 55.50min, 44.75min and 71min. respectively. The *Litsea monopetala* extract displayed significant antioxidant activity which was 20.75 (mg of GAE / gm) of extracts. The activities observed could be attributed to presence of some of the phytochemicals which have been related with cytotoxic, thrombolytic, anthelmintic and antioxidant property.

Keywords: Litsea monopetala; Cytotoxic; Thrombolytic; Anthelmintic and antioxidant activity

1. Introduction

Plant sources contains large amount of bioactive compounds that is beneficial for health as well as provide nutrition to human. For thousands of years people are using plant sources for treating human diseases. Different study suggests that taking diet filled with vegetables and fruits are beneficial for health [1]. According to world health organization 80 percent of world population relies on traditional therapies which are mainly produced from plant extracts or their active components [2.3]. Approximately 10,000 to 15,000 of world's plants have been enumerated for medicinal value but in western medicine only 150-200 plants are in use [4]. Ninety percent of the medicinal plant are collected from wild source and its number is about 722 though South Asian Subcontinent carry only 2000 medicinal plant [5,6].

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Production of ROS and free radicals is caused due to aerobic metabolism and it is must for normal functioning of the human body [7]. From study it is found that excess level of free radical cause several diseases including: cancer, diabetes, cardiovascular and neurodegenerative diseases [8]. Plant extracts contain a large amount of Butylated hydroxyl anisole (BHA), Butylated hydroxyl toluene (BHT), Propyl Gallate (PG) and Tert-Butyl Hydro Quinone (TBHQ), Ascorbic acid (vitamin-c) that is operative against free radical and for cell survival [9,10]. Antioxidants, such as glutathione, vitamin C, vitamin A, vitamin E, total phenolic content are the major antioxidant for animal and plants [11]. Many studies suggest that plant encompasses many pharmacologically active compounds with limited toxicity to normal cell [12].

Cardiovascular disease (CVD) is a heart or blood vessel disease which may arise due to blood clot (thrombus) formation. It is responsible for 80% of CVD deaths in males and 75% of CVD deaths in females all over the world [12]. Myocardial infarction, anorexia, hypertension, stroke etc. occurs as a result of thrombus development. Reduction of blood supply to the liver also enhances due to it. Now-a- days plant sources for antiplatelet, anticoagulant, antithrombotic and thrombolytic activity are the major concern of research for the researcher. Healthcare providers are using tissue plasminogen activator (t-PA), Urokinase (UK), streptokinase (SK) etc. for clot lysis but sometimes they cause hemorrhage, anaphylactic reaction etc. [13,14]. Herbal products are considered safer since ancient times and some studies show that they can exert momentous thrombolytic activity [15,16]. Formerly it is testified about phytochemistry of herbs and their anti-thrombotic activity [17,18].

Helminths infections are the common among a large proportion of the world's population. According to World Health Organization 2 million people are diseased by helminths and 100% school aged children become affected by these worms [19]. Nematodes (round worms), the major phyla of helminths that mostly cause the intestinal infection, onchocerciasis and lymphatic filariasis are the result of filarial worms [20,21]. Different parts of the body are targeted by worms such as *Ascaris*, hookworms, *Trichuris, Enterobius, Strongyloides*, and tapeworms. In developing countries, people of the remote areas specially rural areas are affected mostly [20,22].

Litsea monopetala (Roxb.) Pers. sometimes known as *Tetranthera monopetala* is under the family Lauraceae, known as meda in Bengali and Hindi language. It is mainly found in evergreen forest in Nepal. But outside Nepal it covers from Kumaon to Sikkim, Bangladesh, Burma and southwest China [23,24]. In Bangladesh this plant is widely distributed in Chittagong hill tracts, Sylhet and Sal forests of Gazipur, Madhupur, Dinajpur. It is also found throughout the villages of Bangladesh. It is a medium sized tree having up to 18 meters height with a diameter of 60 cm, leaves are 7.5-23 cm long, elliptic-oblong, usually rounded at both ends, pubescent beneath [25]. The bark of this plant has traditional medicinal use as nerves and bone tonic, stomachache, stimulant, analgesic and antiseptic. Traditionally water extract of bark are used with sugar to treat diarrhea and dysentery in Pakistan and India. Pain arising from blows or bruises or from hard work may relief by the use of powder of bark [26]. Roots are also applicable for the management of pain, bruises and contusions as herbal medicine [27]. The leaves possesses several phytochemicals namely alkaloids, carbohydrates, tannins, flavonoids, steroids etc. having antihyperglycemic, antimicrobial, antidiarrheal and anti-inflammatory activities [26].

The present study was conducted to estimate the thrombolytic, cytotoxic, anthelmintic and antioxidant activity of crude methanolic extracts of leaves of *Litsea monopetala*.

2. Material and methods

2.1 Chemicals

All of the chemicals used in this study were analytical grade.

2.2 Collection and proper identification of plants sample

The plant was collected from the village of Laxmipur district, Bangladesh. It was first identified by the botany department of Noakhali Government University College, Maijdee, Noakhali. Then taxonomical identification was done by Bangladesh National Herbarium Mirpur, Dhaka and 45413 were given as accession number.

2.3 Drying and grinding of plant materials

The collected plant parts (leaves) were separated from undesirable materials or plants or plant parts and washed with water to eradicate adhering dirt. They were sun-dried for one week and then dried in mechanical dryer at $50 - 60^{\circ}$ C.

The plant parts were ground into a coarse powder by mechanical grinder, was stored in an airtight container and kept in a cool, dark and dry place until analysis commenced.

2.4 Extraction of Plant materials

Powered material having a weight of 400 gm ware taken in a clean, flat-bottomed colored glass container and drenched in 2100 ml of methanol at 25° C. To resist the entrance of air the container with its contents was closed properly and preserved for 7 days accompanying occasional shaking and stirring to get better extraction. Extract was filtered through cotton by decantation and finally through Whatman No. 1 filter paper. Final filtrates was concentrated at 40°C by a rotary evaporator [28]. It rendered a gummy concentrate of greenish black color. The gummy concentrate was designated as crude extract of methanol.

2.5 Cytotoxic activity

Simple zoological organism Artemia salina (brine shrimp eggs) as a convenient means was used to determine the cytotoxic activity which is known as brine shrimp lethality bioassay. The brine shrimp eggs were allowed to hatch for two days in artificial sea water (3.8% NaCl solution) and to be matured as nauplii [29,30]. Four (4mg) of sample was dissolved in 200 μ l of DMSO and diluted as 400, 200, 100, 50, 25, 12.5, 6.25, 3.125, 1.5625, 0.78125 μ g/ml by serial dilution in each vial containing 5 ml of saline water (3.8% NaCl solution). 100 μ l DMSO diluted to 5 ml of saline water was used as a negative control. Standard vincristine sulphate was used as positive control. The matured nauplii were inoculated to all experimental and control vials. After 24 hours, each vial was inspected to count lived nauplii using a magnifying glass. Obtained data for each concentration were used to calculate the percent mortality by the following equation

% mortality = (no. of dead nauplii/initial no. of live nauplii) \times 100

Where initial number of live nauplii is 10. The median lethal concentration (LC_{50}) was determined from the graph plotting log of concentration versus percent mortality.

2.6 Thrombolytic activity

In vitro thrombolytic activity of Litsea monopetala was performed by clot lysis method using methanolic extract [31]. A standard clot lysis agent known as streptokinase (SK) was used as a positive control and normal saline (0.9% NaCl solution) was used as a negative control. To a commercially available lyophilised streptokinase vial (S-kinase, Popular Pharmaceuticals Ltd, and Bangladesh) of 15, 00,000 IU. 5 ml phosphate buffered saline (PBS) was added and mixed properly. This suspension was used as a stock solution and diluted to 30000IU and 15000IU conc. which was used as the reference standard for thrombolytic activity. 600 mg of crude methanolic extract of leaves of the Litsea monopetala was dissolved in 0.9% NaCl solution to get a concentration 20 mg/ml. The prepared stock solution was used to make different concentrations of extract in isotonic saline: 2.5, 5, 10 and 20 mg/mL. Venous blood (10 ml) without a history of oral contraceptive or anticoagulant therapy was drawn from healthy human volunteer (n=10) and was transferred to different pre weighed apperdorf tube (0.5 ml/tube) and incubated at 37°C for 45 minutes for clot formation. After clot formation serum was removed without agitating the clot and each tube with clot was weighed again to define the clot weight (clot weight = weight of clot containing tube – weight of tube alone). Different concentrations of the plant extract, 2.5 mg/mL (n = 10), 5 mg/mL (n = 10), 10 mg/mL (n = 10) and 20 mg/mL (n = 10) about 500 μ l was added to each eppendorf tube containing pre weighed clot. As a positive control 500 µl streptokinase (300000 IU and 15000 IU) was used and 500 µl saline water was used as a negative control. All the tubes were then incubated at 37 °C for 90 min and observed for clot lysis. After incubation fluid produced was removed and weighed to observe the difference in weight after clot distraction. Difference obtained in weight taken before and after clot lysis was expressed as percentage of clot lysis [32].

2.7 Anthelmintic test

Adult earthworms were used to study anthelmintic activity due to resemblance of them with the intestinal roundworm parasites of human being and availability [32.33]. The earthworms belonging to species *Pheritima posthuma* (Annelida), about 3-5 cm in length and 0.1- 0.2 cm in width weighing about 0.8-3.04 g, were collected from the moist soil of Noakhali Science and Technology University, Sonapur, Noakhali and thoroughly washed with saline water. Methanolic extracts of leaves of *Litsea monopetala* were used to prepare 10, 20, 40, 60, 80 mg/ml concentration as test sample. Piperazine citrate (10 mg/ml) was used as reference standard solution and saline water for control study. Four earthworms were used in each test sample concentration, reference and control solution to observe the physical change of them and counted their paralysis time and death time.

2.8 Antioxidant activity

In-vitro antioxidant activity of *Litsea monopetala* extract was determined by total phenolic content determination using Folin-Ciocalteu reagent as oxidizing agent and gallic acid as standard [33,34]. Gallic acid solution were prepared having a concentration ranging from 100 μ g / ml to 0 μ g / ml. 2.5 ml of Folin-Ciocalteu reagent (diluted 10 times with water) and 2.0 ml of Na₂CO₃ (7.5 % w/v) solution was added to 0.5 ml of gallic acid solution. The mixture was incubated for 20 minutes and absorbance was measured at 760 nm to prepare a standard curve. To 2 mg / ml extract concentration 2.5 ml of Folin-Ciocalteu reagent (diluted 10 times with water) and 2.0 ml of Na₂CO₃ (7.5 % w/v) solution was added, incubated for 20 minutes and absorbance was measured at 760 nm. Standard curve prepared from gallic acid solution with different concentration and the total phenol content of the sample was measured as mg of GAE (gallic acid equivalent) / gm of the extract.

2.9 Statistical Analysis

The data are expressed as the mean ± SEM analyzed by one-way analysis of variance (ANOVA) and Dunnett's t-test was used as the test of significance. P value <0.05 was considered as the minimum level of significance. All statistical tests were carried out using SPSS (version 16) statistical software.

3. Results and discussion

3.1 Brine shrimp lethality bioassay

The brine shrimp test (BST) represents a rapid, inexpensive and simple bioassay for testing plant extract lethality which in most cases correlates reasonably well with cytotoxic and anti-tumor properties. The summary of the result was given below (Table 3). Here the LC₅₀ for standard Vincristine Sulphate is 0.839 (μ g/ml) and for methanolic extract of *Litsea monopetala* is 41.05 (μ g/ml) (Table 1, 2).

Methanol Extract				Vincristine Sulphate			
Conc. (C) (µg/ml)	Log C	% Mortality	LC ₅₀ (µg/ml) Based on Log C	Conc. (C) (µg/ml)	Log C	% Mortality	LC ₅₀ (μg/ml) Based on Log C
400	2.602059991	100		40	1.602059991	100	
200	2.301029996	90		20	1.301029996	90	
100	2.00000000	60		10	1.00000000	90	
50	1.698970004	50		5	0.698970004	80	
25	1.397940009	20	44.05	2.5	0.397940009	70	0.839
12.5	1.096910013	20	41.05	1.25	0.096910013	70	
6.25	0.795880017	10		.625	-0.20411998	50	
3.125	0.494850022	10		.3125	-0.50514997	30	
1.5625	0.193820026	00		.15625	-0.80617997	20	
.78125	-0.10720997	00		.07813	-1.10720997	10	

Table 1 Effect of Methanolic Extract of Litsea monopetala Leaves on Brine Shrimp Nauplii

Table 2 Results of the test sample of Litsea monopetala leaves

Sample	LC ₅₀ (µg/ml)	Regression Equation	R ²
Vincristine Sulphate (Positive control)	0.839	y=34.02x+52.58	0.952
Methanol Extract	41.05	Y=38.25-11.71	0.893

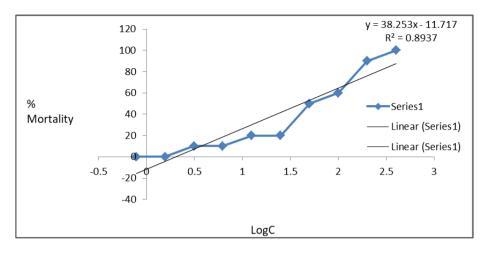


Figure 1 Effect of Methanol Extract on Brine Shrimp Nauplii

3.2 In-Vitro Thrombolytic Activity

Table 3 Effect of methanolic crude extracts of stem of *Litsea monopetala* on blood clot lysis of human blood *in vitro*(mean± SEM)

Concentrations of plant extracts, control and standard	n	Mean % of Blood clot lysis		
0.9% NaCl solutions		5.3540±1.01*		
Streptokinase (30,000 I.U.)		47.2189±1.15*		
Streptokinase (15,000 I.U.)		24.7321±1.12*		
Leaves extract 2.5 mg/mL		9.5231±1.24		
Leaves extract 5 mg/mL	10	9.4987±1.20		
Leaves extract 10 mg/mL		13.6436±1.22		
Leaves extract 20 mg/mL		17.5013±1.29*		
* Determines significance level				

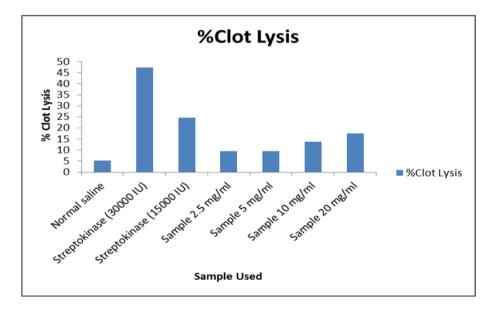


Figure 2 Clot lysis of blood samples of normal subjects by different concentrations of crude methanolic extracts of leaves of *L. monopetala*

The clots were treated by four different concentrations of leaves extracts i.e., 2.5, 5, 10 and 20 mg/ml and clot lysis % was 9.52, 9.49, 13.64 and 17.50 % respectively (Table 5). Standard streptokinase 30000 IU and 15000 IU evoked a significant (p<0.001) clot lysis 47.22 and 24.73 % respectively (Table 3). This effect showed a dose related trend.

3.3 Anthelmintic test

From the study it was observed that the extract of *Litsea monopetala* showed not only paralysis but also death of earthworms. Whereas methanol extract at different concentrations showed paralysis at 76.75 min, 60 min, 51.75 min, 44.5 min and 64.5 min and death at 90.50 min, 63.75 min, 55.50 min, 44.75 min and 71 min. for 10 mg, 20 mg, 40 mg, 60 mg and 80 mg respectively. The standard drug piperazine at 10 mg/ml concentration shows paralysis at 56.2 and death at 71min of earthworms respectively.

Test Substance	Concentration (mg/ml)	Time taken for paralysis (Mean± SEM)	Time taken for death (Mean± SEM)	
Control (Distilled water)	10	0.00	0.00	
Standard (Piperazine)	10	56.2±.20000	71±.244	
Methanolic extract	10	76.75±1.65	90.50±0.64	
	20	60.00±0.91	63.75±0.85	
	40	51.75±0.62	55.50±1.25	
	60	44.50±2.10	44.75±1.49	
	80	64.50±0.20	71±0.24	

Table 4 Paralysis and death time of *Pheritima posthuma*

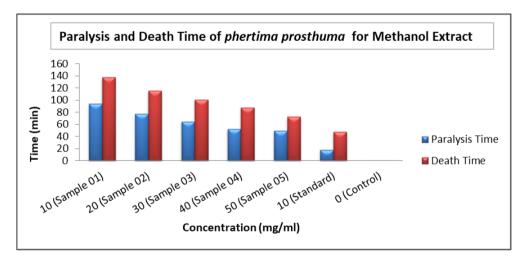


Figure 3 Paralysis and Death Time of phertima prosthuma for Methanol Extract

3.4 Total phenolic content determination

Total phenolic content of the samples are expressed as mg of GAE (gallic acid equivalent)/ gm which was determined by Folin-Ciocalteu reagent and gallic acid as standard. The amount of total phenolic content of methanol extract of plant of *Litsea monopetala* is 20.75 mg of GAE / gm of extract (Table 5).

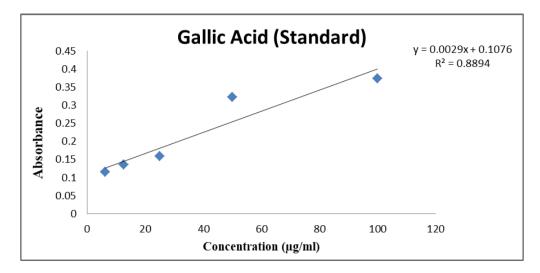


Figure 4 Total Phenolic Content of Gallic Acid (Standard)

Table 5 Total Phenolic Content of Gallic Acid (Standard)

Conc. of the Standard (μ g/ml)	Avg. Absorbance at 760 nm	Regression line	R ²
100	0.373		
50	0.323		
25	0.159	y = 0.002x + 0.107	0.889
12.5	0.135		
6.5	0.115		

Table 6 Total Phenolic Content of Litsea monopetala

Extract	Absorbance of the sample	Total Phenolic Content (mg of GAE / gm) of Extracts
Methanol Extract	0.190	20.75

4. Discussion

For drug development cytotoxicity should be taken into consideration. The leaves extract of *Litsea monopetala* shows minimal amount of cytotoxicity which is 41.05 (μ g/ml) because the biological activities of plants may be due to the presence of diverse group of chemical compounds like glycosides, alkaloids, flavonoids and saponis [30].

Most thrombolytic agent activates the enzyme plasminogen which clears the cross-linked fibrin mesh, makes the blood soluble and refurbishes blood flow over occluded blood vessels. That's why thrombolytic agents are beneficial for the treatment of myocardial infarction, thromboembolic strokes, deep vein thrombosis and PE to clear a blocked artery and avoid permanent damage to the perfused tissue (e.g. myocardium, brain, and leg). All four concentrations of crude methanolic extracts of stem of *Litsea monopetala* induced significant (p < 0.001) clot lysis activity in vitro, compared to control which shows a dose-related trend ($r^2 = 0.7565$; p<0.001). Study shows that *Litsea monopetala* contains phytosterols which is responsible for clot lysis [33].

From the study it was observed that the methanolic extracts of Litsea monopetala showed not only paralysis but also death of earthworms which shows a dose related trend. Preliminary phytochemical screening of methanol extract showed the presence of saponins, tannins and alkaloids which interfere with helmintic parasites [20,33]. Therefore, the anthelmintic activity of methanol extract as described herein against earthworms suggests that it could be effective against parasitic infections of humans.

Phenolic compounds allow them to act as antioxidants due to their redox properties. The total phenolic concentration could be used as a basis for rapid screening of antioxidant activity as their free radical scavenging ability is facilitated by their hydroxyl groups [33,34]. The amount of total phenolic content was significant in compared standard Gallic acid. Hence this study was conducted by crude extract, further advanced studies should be carried out for compound isolation and it is necessary to observe which compounds are actually responsible for specific effects.

5. Conclusion

Results of our study suggest the great value of the species *L. monopetala* for use in pharmacy and phytotherapy. Based on this information, it could be concluded that this plant is natural sources of antioxidant substances of high importance. The methanol extract shows a limited amount of cytotoxic activity. In case of thrombolytic and anthelmintic activity the extract was an appreciable effect. This is only a preliminary study. Further phytochemical analysis is required to isolate the elements of the plant to ensure the use in human health remedy like Cancer, Cardio vascular disease, Ageing problem, anthelmintic effect etc.

Compliance with ethical standards

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Disclosure of conflict of interest

There is not any conflict of interest.

References

- [1] Montuschi P, Barnes JP, Roberts JL. (2004). Isoprostanes: markers and mediators of oxidative stress. The FASEB Journal, 18(15), 1791-1800.
- [2] Roja G, Rao C. (2000). Anticancer compounds from tissue cultures of medicinal plant. Journal of Herbs, Spices and Medicinal Plants, 7, 71-102.
- [3] Bekker J, Ploem S, de Jong KP. (2009). Early hepatic artery thrombosis after liver transplantation: A systematic review of the incidence, outcome and risk factors. Am J Transplant, 9(4), 746-757.
- [4] Anwar SM, Khan IN, Sarkar MM, Barua S, Kamal ATMM, Hosen MZ. (2011). Thrombolytic & cytotoxic effect of different herbal extracts. IJPSR, 2(12), 3118-3121.
- [5] Sherwani SK, Bashir A, Haider SS, Shah MA, Kazmi SU. (2013). Thrombolytic potential of aqueous and methanolic crude extracts of *Camellia sinensis* (Green Tea): *In vitro* study. Journal of Pharmacognosy and Phytochemistry, 2(1), 125-129.
- [6] Raju GS, Moghal MMR, Dewan SMR, Amin MN, Billah MM. (2013). Characterization of phytoconstituents and evaluation of total phenolic content, anthelmintic, and antimicrobial activities of *Solanum violaceum* Ortega. Avicenna Journal of Phytomedicine, 3(4), 313-320.
- [7] Baul S, Amin MN, Hussain MS, Mukul MEH, Millat MS, Rashed MSU, et al. Phytochemical Nature and Pharmacological Evaluation of Chloroform Extract of Pandanus fascicularis L. (Fruits) An in vivo Study. Journal of Bioanalysis & Biomedicine. 2017; 9(4): 223-228.
- [8] Dewan SMR, Amin MN, Adnan T, Uddin SMN, ShahidUd-Daula AFM, Sarwar G. (2013). Investigation of analgesic potential and in vitro antioxidant activity of two plants of Asteraceae family growing in Bangladesh. Journal of Pharmacy Research, 6(6), 599-603.
- [9] Uddin SMN, Amin MN, Shahid-Ud-Daula AFM, Hossain H, Haque MM, Rahman MS. (2014). Phytochemical screening and study of antioxidant and analgesic potentials of ethanolic extract of *Stephania japonica* Linn. Journal of Medicinal Plant Resrearch, 8(37), 1127-1133.
- [10] Amin MN, Dewan SMR, Noor W, Shahid-Ud-Daula AFM. (2013). Characterization of chemical groups and determination of total phenolic content and in-vitro antioxidant Activities of ethanolic extract of *Ocimum sanctum* leaves growing in Bangladesh. European Journal of Experimental Biology, 3(1), 449-454.

- [11] Amin MN, Banik S, Ibrahim M, Moghal MMR, Majumder MS, Siddika R. (2015). A Study on *Ardisia solanacea* for Evaluation of Phytochemical and Pharmacological Properties. International Journal of Pharmacognosy and Phytochemical Research, 7(1), 8-15.
- [12] Tanna MTH, Amin MN, Ibrahim M, Mukul MEH, Kabir A. (2016). Evaluation of antioxidants, membrane stabilizing, cytotoxic and anthelmintic activity with phytochemical screening of *Chromolaena odorata*: A medicinal shrub. International Journal of Pharmacy, 6(1), 53-61.
- [13] Amin MN, Siddiqui SA, Uddin MG, Ibrahim M, Uddin SM. (2020). Increased Oxidative Stress, Altered Trace Elements, and Macro-Minerals Are Associated with Female Obesity. Biol Trace Elem Res., 197, 384–393.
- [14] Amin MN, Hussain MS, Sarwar MS, Rahman Moghal MM, Das A, Hossain MZ. (2019). How the association between obesity and inflammation may lead to insulin resistance and cancer. Diabetes Metab Syndr., 13(2), 1213-1224.
- [15] Islam T, Hussain MS, Amin MN, Tuhin AM. (2018). Evaluation of psychopharmacological and neurosafety profile of Swas Kas Chintamani Ras (SKC) in Swiss-Webster mice. Avicenna J Phytomedicine, 8(1), 85-95.
- [16] Amin MN, Liza KF, Sarwar MS, Ahmed J, Adnan MT, Chowdhury MI. (2015). Effect of lipid peroxidation, antioxidants, macro minerals and trace elements on eczema. Arch Dermatol Res., 307(7), 617-23.
- [17] Ghosh A, Banik S, Amin MN, Ahmed J. (2018). Evaluation of antinociceptive, antihyperglycemic, and membrane stabilizing activities of *Garcinia lancifolia* Roxb. J Tradit Complement Med., 8(2), 303-307.
- [18] Muktadir MHA, Islam MA, Amin MN, Ghosh S, Siddiqui SA, Debnath D. (2019). Nutrition transition Pattern IV: Leads Bangladeshi youth to the increasing prevalence of overweight and obesity. Diabetes Metab Syndr., 13(3), 1943-1947.
- [19] Islam MA, Amin MN, Siddiqui SA, Hossain MP, Sultana F, Kabir MR. (2019). Trans fatty acids and lipid profile: A serious risk factor to cardiovascular disease, cancer and diabetes. Diabetes Metab Syndr., 13(2), 1643-1647.
- [20] Su JX, Wang W, Zhang LB, Chen ZD. (2006). Phylogenetic placement of two enigmatic genera, Borthwickia and Stixis, based on molecular and pollen data, and the description of a new family of Brassicales, Borthwickiaceae. Taxon, 61(3), 601-611.
- [21] Reddy ARK, Grace JR. (2016). *In vitro* Evaluation of Antioxidant Activity of Brugeiera Gymnorrhiza and Aegialitis Rotundifolia. Med Aromat Plants, 5, 231.
- [22] Sikder MAA, Kaisar MA, Rashid MA, Millat MS, Sultana A. (2012). *In vitro* membrane stabilizing activity, total phenolic content, cytotoxic, thrombolytic and antimicrobial activities of *Calliandrasuri namensis* (Wall.). J. Pharmacog. Phytochem., 1(3), 45-50.
- [23] Škerget M, Kotnik P, Hadolin M, Hraš AR, Simonič M, Knez Ž. (2005). Phenols. proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food chemistry, 89(2), 191-198.
- [24] Rahaman MZ, Akhter S, Islam MR, Begum S, Mondal KK, Mottakin M. (2020). Assessment of thrombolytic, antioxidant and analgesic properties of a medicinal plant of Asteraceae family growing in Bangladesh. Discovery Phytomedicine, 7(1), 47-52.
- [25] McChesney JD, Venkataraman SK, Henry JT. (2015). Plant natural products: back to the future or into extinction?. Phytochemistry, 68(14), 2015-22.
- [26] Cartea ME, Francisco M, Soengas P, Velasco P. (2011). Phenolic compounds in Brassica vegetables. Molecules, 16(1), 251-280.
- [27] Rahman AHMM, Sultana N, Islam AKMR, Zaman ATMN. (2013). Study of medical ethno-botany at the village Genda under Savar Upazilla of District Dhaka, Bangladesh. International Journal of Medicinal Plants Research, 2(1), 18-31.
- [28] Giuseppina B, Cristiana L, Guido L, Piero C, Antonio LA, Daniele R. (2004). Therapeutic effect of diagnostic ultrasound on enzymatic thrombolysis. An *in vitro* study on blood of normal subjects and patients with coronary artery disease, Journal of Thrombosis and Haemostasis, 91, 1078-1083.
- [29] Steinmann P, Keiser J, Bos R, Tanner M, Utzinger J. (2006). Schistosomiasis and water resources development: Systemic review, meta-analysis, and estimates of people at risk. Lancet Infect. Dis., 6, 411-425.
- [30] Hasan H, Azad MSL, Islam MZ, Rahman SM, Islam MR, Rahman S, Rahmatullah M. (2014). Antihyperglycemic activity of methanolic extract of *Litsea monopetala* (Roxb.) Pers. leaves. Adv Nat Appl Sci., 8, 51-55.

- [31] Hossain S, Kader G, Nikkon F, Yeasmin T. (2012). Cytotoxicity of the rhizomes of medicinal plants. Asian Pac J Trop Biomed., 2(2), 125–127.
- [32] Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Daginawala HF. (2006). Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs. Thromb J., 4(14), 1–4.
- [33] Meyer BN, Ferrigni NR, Putnam JE, Jacobsen JE, Nichols DE, McLaughlin JL. (1982). Brine shrimp: A convenient general bioassay for active plants constituents. J Med Plant Res., 45, 31–34.
- [34] Vital PG, Rivera WL. (2011). Antimicrobial activity, cytotoxicity, and phytochemical screening of *Voacanga globosa* (Blanco) Merr. leaf extract (Apocynaceae). Asian Pac J Trop Med., 4(10), 824–828.